Experimental Goettingen Minipig and Beagle Dog as Two Species Used in Bioequivalence Studies for Clinical Pharmacology (5-Aminosalicylic Acid and Atenolol as Model Drugs)

J. KVĚTINA, Z. SVOBODA, M. NOBILIS, J. PASTERA AND P. ANZENBACHER

Institute of Experimental Biopharmaceutics, Joint Laboratory of Czech Academy of Sciences and PRO.MED.CS, Hradec Králové, Czech Republic

Abstract. Due to proven similarities in biotransformation between man and minipig, minipig seems to be the experimental animal of choice for preclinical pharmacokinetic studies when an experiment with a drug exhibiting a great first pass bioelimination (like 5-aminosalicylic acid) is to be realised. On the other hand, both minipig and dog may be suitable species for a pharmacokinetic study with a drug characterized by a small extent of first pass biotransformation (like atenolo).

Key words: Pharmacokinetics — Atenolol — 5-aminosalicylic acid — Man — Dog — Minipig

Introduction

One of the limiting factors in the application of preclinical pharmacokinetic data to clinical studies with human volunteers is the choice of an adequate experimental animal species. For the solid p.o. dosage pharmaceutical forms, the choice is rather narrow as only the omnivorous animals as dogs, pigs and occasionally primates (macaca species) are available. There is evidence (Lázníček and Květina 1988) that the inter-species differences are based:

- on quantitative aspects such as the extent of drug binding, transport across the biological membranes,

- on qualitative differences, e.g. drug biotransformation.

The aim of this study is to compare experimental animals according to the character of the pharmacokinetic data for studied drugs. The postulate was tested experimentally with two model drugs:

- atenolol: a beta-blocking agent (Wood 1984) with a small extent of first pass biotransformation of the parent drug in man (less than 15 %, Lennard et al. 1986)

Correspondence to: Prof. Dr. Jaroslav Květina, DrSc., ÚEBF, 500 02 Hradec Králové, Heyrovského 1207, Czech Republic. E-mail: uebf@hk.cesnet.cz

with the following kinetic parameters (Riddell et al. 1987) t_{max} = 90 (± 30) min, t_{1/2} = 6–9 h,

– 5-aminosalicylic acid: an antiphlogistic agent used in the treatment of ulcerative colitis (Greenfield et al. 1993) which is rapidly biotransformed in man to N-acetyl-metabolite (Klotz 1985): t_{max} of the parent drug = 105 (±35) min, urinary excretion of the parent drug = 80%; t_{max} of N-acetyl-5-ASA = 39 (±12) min, urinary excretion of the metabolite = 20%.

Abbreviations: ATE, atenolol, 5-ASA, 5-aminosalicylic acid; N-acetyl-5-ASA, N-acetyl-metabolite of 5-aminosalicylic acid; AUC, area under the curve, c_{max} , maximal plasma concentration, t_{max} , time at which maximal plasma concentration was reached; $t_{1/2}$, half-time of elimination.

Materials and Methods

Materials

Both compounds tested, ATE and 5-ASA, were applied as commercially available solid dosage forms ATE as Atenormal tab 100 mg (PRO MED CS Praha as, Czech Republic) and 5-ASA as Salozinal tab 500 mg (PRO MED CS Praha as, Czech Republic) Both dosage forms were administered orally as a single dose All chemicals used to preparate solutions were obtained from Sigma Aldrich (Praha, Czech Republic) and were of reagent grade

Experimental design

Experimental animals fasted overnight and for 2 h after oral drug administration Blood samples were obtained before drug intake and at various intervals after each administration (at 15, 30, 60 min, 2, 4, 8, 12, 16, 20, 24 h, in the case of man also 48 h) Five Goettingen minipigs (average weight 245 ± 15 kg, males) were used, blood samples were obtained from incannulated jugular vein From five Beagle dogs (average weight 165 ± 23 kg, males), blood samples were obtained from veins at the surface of the legs (v cephalica antebracchii and v metatarsea dorsalis) Human subjects in the study were healthy middle-aged volunteers (12 in the case of ATE, 24 for 5-ASA) All experiments were approved by the corresponding ethical committees

Assays

ATE HPLC determination using a 150×4 mm column packed with Lichrospher CN and with an acetonitrile-phosphate buffer as the mobile phase was performed using metoprolol as internal standard. The accuracy and precision of atenolol determination were found to range from 88.6 % to 96.8 % and from 1.0 % to 7.9 %, respectively

5-ASA Plasma levels of both the parent compound and the metabolite were determined by the reverse-phase HPLC method with a 250×4 mm column packed with Lichrospher RP-18e stationary phase The analytes were eluted with acetonitrile / octanesulphonic acid buffer pH 2.8 with simultaneous UV and fluoroscence detection N-acetyl-4-ASA was the internal standard The accuracy and precision of the N-acetyl-5-ASA determination were found to range from 92.9 % to 107.4 % and from 0.7 % to 8.6 %, respectively In the case of 5-ASA, the accuracy and precision were found to range from 87.3 % to 100.9 % and from 1.1 % to 3.2 %, respectively

In both cases an HPLC system (ThermoSeparation Products, USA) was used consisting of a quaternary pump, an autosampler and a SpectraFocus UV/VIS detector

Results

Plasma levels of both compounds (ATE and 5-ASA) as well as of the 5-ASA metabolite (N-acetyl-5-ASA) were determined. From the plasma concentration-tume curves, three pharmacokinetic parameters were determined: AUC, c_{max} , t_{max} . The parameters were calculated according to standard mathematical procedures (AUC, trapezoidal method; c_{max} and t_{max} were taken from experimental data).

For ATE the pharmacokinetic parameters were similar in all the three species examined (man, minipig, dog) (Table 1). This follows also from the course of the experimental curves (Fig. 1) when the dose is expressed relative to the dose and body weight.

In the case of 5-ASA, the plasma levels in man (Fig. 2) and minipig (Fig. 3) were lower than those of N-acetyl-5-ASA. In the dog, no formation of N-acetyl-5-ASA was observed (Fig. 4). Pharmacokinetic parameters of both the parent compound 5-ASA and the metabolite were similar in man and minipig when the dose was expressed relative to the dose and body weight (Table 2).

Table 1. Pharmacokinetic parameters of atenolol in man, minipig and dog (expressed relatively dose to body weight)

| | man | minipig | dog |
|-----------------------------|-------------------|--------------------|------------------|
| AUC [ng h/ml/D] | 3934 3±1376 1 | $2601\ 1\pm509\ 2$ | 3226 4±623 9 |
| $c_{\rm max} [\rm ng/ml/D]$ | 384.8 ± 168.4 | $331\ 1\pm 89\ 9$ | $443\ 5\pm74\ 2$ |
| $t_{\rm max}$ [h] | 2.6 ± 1.1 | $1.5{\pm}0.6$ | 1.5 ± 0.7 |

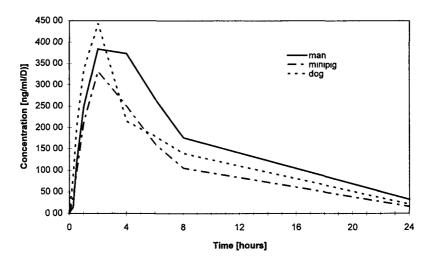


Figure 1. Plasma levels of atenolol after oral administration (100mg) to man, minipig and dog

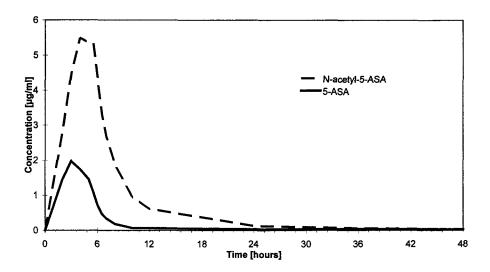


Figure 2. Levels of 5-aminosalicylic acid and N-acetyl-5-aminosalicylic acid in human plasma after oral administration of 5-aminosalicylic acid (500 mg).

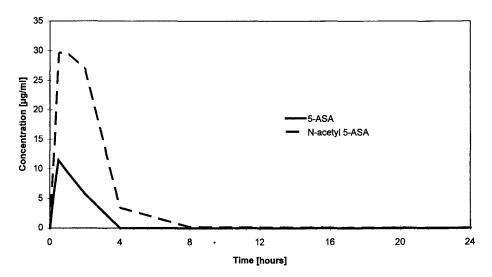


Figure 3. Levels of 5-aminosalicylic acid and N-acetyl-5-aminosalicylic acid in minipig plasma after oral administration of 5-aminosalicylic acid (500 mg).

Discussion

Man and minipig are known to exhibit nearly similar physiology of the intestinal and kidney tracts (Leucht et al. 1982; Swindle and Smith 1998). Moreover, there

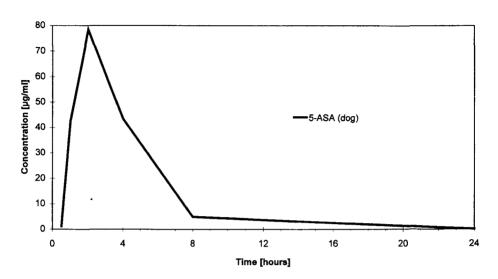


Figure 4. Levels of 5-aminosalicylic acid and N-acetyl-5-aminosalicylic acid in dog plasma after oral administration of 5-aminosalicylic acid (500 mg)

| | man | minipig | \log |
|---|--|---|--|
| -ASA | | | |
| AUC [µg h/ml/D] _{max} [µg/ml/D] _{max} [h] | $\begin{array}{c} 1 \ 4 \pm 0 \ 8 \\ 0 \ 4 \pm 0 \ 2 \\ 3 \ 5 \pm 1 \ 1 \end{array}$ | $\begin{array}{c} 1 \ 3\pm 0 \ 5 \\ 0 \ 9\pm 0 \ 6 \\ 1 \ 4\pm 0 \ 8 \end{array}$ | $egin{array}{c} 11\ 2\pm 3\ 1\ 3\ 2\pm 1\ 8\ 1\ 8\pm 0\ 5 \end{array}$ |
| -acetyl-5-ASA | | | |
| UC [μg h/ml/D] _{max} [μg/ml/D] _{max} [h] | $\begin{array}{c} 4 \ 8 \pm 1 \ 1 \\ 1 \ 0 \pm 0 \ 4 \\ 4 \ 3 \pm 1 \ 2 \end{array}$ | $5\ 4\pm1\ 5\ 2\ 4\pm0\ 9\ 1\ 4\pm0\ 8$ | |

 Table 2. Pharmacokinetic parameters of 5-ASA and N-acetyl-5-ASA in man, minipig and dog (expressed relatively dose to body weight)

is a considerable similarity in the activities of the enzymes of biotransformation It has been shown recently that the enzymes of "Phase I of biotransformation", mainly the cytochromes P-450, possess similar properties in the minipig and in man (Anzenbacher et al. 1998; Witkamp and Monshouwer 1998). The similarity of substrate specificities of minipig and human drug-metabolizing systems has been confirmed recently (Anzenbacher et al. 1998; Madden et al. 1998). On the other hand, the dog seems to be, at least in some cases, a less suitable or less versatile experimental animal for studies on drug biotransformation (Chauret et al. 1997; Witkamp and Monshouwer 1998). This is documented in our study by the sole fact that there was no N-acetyl metabolite of 5-ASA found in plasma samples from this experimental animal

The present study represents a contribution to the current discussion on the suitability of various species of experimental animals for modelling of pharmacokinetics in man For drugs with relatively small "first pass" effect of biotransformation, there is a greater choice of suitable animal species for studies of comparative pharmacokinetics (as e.g. in the case of atenolol, where the pharmacokinetics parameters were similar for man, minipig as well as for dog). On the other hand, when the "first pass" effect is more pronounced, the situation is more complicated as the choice of the suitable species much more depends on mechanisms of biotransformation of the given drug and on the enzymes of biotransformation active or available in the particular animal species.

References

- Anzenbacher P, Souček P, Anzenbacherová E, Gut I, Hrubý K, Svoboda Z, Květina J (1998) Presence and activity of cytochrome P450 isoforms in minipig liver microsomes Drug Metab Disposition 26, 56—59
- Chauret N, Gauthier A, Martin J, Nicoll-Griffith D A (1997) In vitro comparison of cytochrome P-450-mediated metabolic activities in human, dog, cat, and horse Drug Metab Disposition **25**, 1130–1136
- Greenfield S M, Punchard N A, Teare J P, Thompson R P H (1993) The mode of action of the aminosalicylates in inflammatory bowel disease Aliment Pharmacol Therapeut 7, 369–383
- Klotz U (1985) Clinical pharmacokinetics of sulphasalazine, its metabolites and other prodrugs of 5-aminosalicylic acid Clin Pharmacokinet 10, 285-302
- Lázníček M, Květina J (1988) The effect of molecular structure on the distribution and elimination of some organic acids Quant Struct Act Relat 7, 234–239
- Lennard M S, Tucker G T, Woods H F (1986) The polymorphic oxidation of betaadrenoreceptor antagonists Clinical pharmacokinetic considerations Clin Pharmacokinet 11, 1--17
- Leucht W, Gregor G, Stier H (1982) Einfuhrung in die Versuchtierkunde Band IV Das Miniaturschwein, Gustav Fischer, Jena
- Madden S, Gentile D M, Grawford G A, Simpson A G, Johnston A M (1998) Cytochrome P-450 activities in hepatic microsomes from the minipig ISSX (Int Soc Study Xenobiotics) Proceedings Vol 13, 75
- Riddell J G, Harron D W G, Shanks R G (1987) Clinical pharmacokinetics of betaadrenoreceptor antagonists Clin Pharmacokinet 12, 305–320
- Swindle M M, Smith A C (1998) Comparative anatomy and physiology of the pig Scand J Lab Anim Sci Suppl 25, 11—22
- Witkamp R F, Monshouwer M (1998) Pharmacokinetics in vivo and in vitro swine Scan J Lab Anim Sci Suppl 25, 45-56
- Wood A J J (1984) Pharmacologic differences between beta blockers Amer Heart J 108, 1070–1077